

Fitting Models of Vulnerability to Toxicity with Generalized Linear Models

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Abstract: People are often exposed to toxic or hazardous (e.g. radioactive radon and lead) elements and rays, without even knowing so. Toxicity often results from an individual's prolonged exposure to toxic substances. A thorough examination of some individuals' blood or urine samples for the quantities of hazardous substances or elements, often gives a multivariate data (i.e. matrix of cases against elements) on toxicity. The pertinent response variable is often binary response (or count data) type and hence the Generalized Linear Models (GLM) of it can be fitted using our proposed techniques. This paper purports to identify models in GLM that can be used to study toxicity when it is 'captured' as count data or Binary Response Variables (BRV). An illustration of how the techniques work is done by using a sample of data on some artisans.

Keywords: GLM, Exploratory Data Analysis (EDA), BRV, Count Data (CD), Toxicity, R

1. Introduction

Pollution happens in various ways; environmental and occupational exposures to pollutants are usually experienced by artisans [4]. Environmental pollution can sometimes be due to human inappropriate activities (e.g. the dumping of toxic waste in residential locations) or natural (e.g. the natural emission of radioactive radon in residential buildings (indoor radon)) [5] [6] [7]. In the former case, the activity can be easily stopped, whilst in the latter case, little or nothing can be done. With respect to occupational exposure; some measures can be put in place to usurp their effects on humans or, at least, reduce the effects to the barest minimum. It is because of occupational exposure that the technologists, and artisans, working in radioactive environments, are strongly advised to display the 'symbol' for radioactivity in a conspicuous location around their laboratories and workshops respectively and to always use protective gadgets and advise their patrons and customers to do the same. Accidental exposure is also possible, it may happen in a mineral mining field or at a nuclear energy station such as the one of Chernobyl and Fukushima [4]. When people are in a polluted environment, they are said to be exposed to dangerous (or toxic) substances (e.g. indoor radon, fungi spores, and lead). Hence toxicity, in an individual, often

results from his/her prolonged exposure to toxic substances. The individual will be 'pronounced' toxic, with respect to the toxic substance, if the estimated quantity of the substance found in the samples (e.g. blood or urine) from his/her body is higher than the quantity that can be tolerated by a human body (i.e. without associating any allied ailments). Upon a thorough examination of some individuals' blood or urine samples for the quantities of 'hazardous' substances or elements, a multivariate data (i.e. matrix of cases against elements) on toxicity will be obtained [3]. With respect to a count data or response variable (y_i , $i = 1, 2, \dots, n$), that is, dichotomous in nature, generalized linear models of toxicity can be fitted [2] [6] [7]. EDA tools are very 'restricted', in usage, and subject to misinterpretations with respect to these two cases (i.e. count data and binary response variables) because the numerical code of each BRV, say, is either zero (0) or one (1).

2. Exploratory Data Analysis (EDA) and Binary Response Variables (BRV)

Any BRV (y) is necessarily dichotomous in nature. That

is, it can have either of the following pairs of responses; yes or no, high or low, tall or short, diseased or not-diseased, alive or dead etc. BRVs are usually coded with 1 or 0 with respect to the analyst's discretion. For instance, an analyst may adopt the following with respect to his/her BRV for a particular work:

$$y_i = \partial_0^1 = \begin{cases} 1, & \text{yes} \\ 0, & \text{no} \end{cases} \quad i = 1, 2, \dots, n \quad (1)$$

Then, the variable $\bar{y} = \frac{\sum_{i=1}^n y_i}{n}$ that is usually known as the

sample's measure of central tendency or mean ($E(y)$) is now simply the 'proportion (p)' of the responses that are coded 1 (i.e. yes). Similarly; the variance ($V(y)$) can be expressed in terms of p, as shown below:

$$V(y) = \frac{\sum_{i=1}^n (y_i - p)^2}{n} = \frac{\sum_{i=1}^n y_i^2}{n} - p^2 = \frac{\sum_{i=1}^n y_i}{n} - p^2 = p(1-p) = pq \quad (2)$$

Where $y^2 = (\partial_0^1)^2 = (\partial_0^1) = y$ and $p + q = 1$. Equations (1) and (2) are strong indications for the Bernoulli ($Ber(p)$) distribution. Because of the relationships existing amongst the; Bernoulli, Binomial, Poisson, Normal (i.e. the Exponential Family of Distributions (EFD)), it is reasonable to model the BRV with the GLM which 'toggles' around EFD easily. The choice of an EDA tool however, may be inappropriate because, they often results in non-informative descriptive or pictorial representations. For example a histogram or a box-plot will contain just two pictorial representations with very 'little' information on y. Also the stem-and-leaf plot often results into descriptive representation having just two lines of zeros and ones. Although cluster analysis still possess 'little' usefulness but they can only be used to 'split' the responses into just two clusters as well. This paper purports to identify models in GLM that can be used to study toxicity when it is 'captured' as count data or BRV.

3. Toxicity and GLM in R

GLM are extensions of traditional regression models that allow the mean to depend on the explanatory variables through a link function (e.g. log, logit, probit, cloglog, identity, sqrt) and the response variable to be any member of a set of distributions called the EFD. Toxicity can be studied through GLM and the R language in two ways; when the 'experimental units' or organisms are monitored to mortality and when 'experimental units' are just 'screened' for vulnerability to toxicity. The R function for fitting a generalized linear model is "*glm()*". There are many methods (or commands) for 'glm objects', they include; "summary",

"coef", "resid", "predict", "anova" and "deviance" [2].

3.1. Assumptions on Variables and General Setup for GLM in R

Throughout, we shall assume that;

1. BRV or count data ($Y (n \times 1)$) and their corresponding multivariate data ($X (n \times m)$) can be represented as below

$$Y = \begin{pmatrix} y_1 \\ \vdots \\ y_n \end{pmatrix}, \quad X = \begin{pmatrix} x_{11} & \dots & x_{1m} \\ \vdots & & \vdots \\ x_{n1} & \dots & x_{nm} \end{pmatrix} \quad (3)$$

2. Y further follows one member of the EFD (e.g. $Y \sim Bin(n, p)$) with some parameterization say, ξ known as the "linear predictor", such that $\xi = \beta X$.

We are required to estimate the parameters ($\beta (m \times 1)$). Now let η and μ denote the natural and mean parameterizations of the pertinent member of the EFD. Then;

3. There is a scale parameter ϕ through which we can estimate over-dispersion. Over-dispersion essentially describes the situation whereby the actual $V(Y_i)$; for some $i = 1, 2, \dots, n$, exceeds the GLM variance $\phi V(\eta_i)$.

4. There is a link function ℓ defined by $\ell(\mu_i) = \xi_i, \quad i = 1, 2, \dots, n$.

5. There is a canonical link function ℓ defined by $\ell(\mu_i) = \eta_i, \quad i = 1, 2, \dots, n$.

3.2. When the Organisms are Monitored to Mortality

When the "experimental units" are monitored to mortality, modelling the toxicity with GLM assumes that the toxicity increases with the increase in; exposure of the units, dose of the toxic element until when the units get exterminated. The following example illustrates the how the technique works. The data was obtained when groups of 20 male and female moths were exposed to six "increasing" levels of a "pyrethroid" in order to "capture" its toxicity to tobacco budworm. The technique is contained in the following R commands:

```
> library("rJava")
> library("glmulti")
> ldose<-rep(0:5,2)
> numdead<-c(1,4,9,13,18,20,0,2,6,10,12,16)
> sex<-factor(rep(c("M","F"), rep(6,2)))
> SF<-cbind(numdead, numalive=20-numdead)
> budworm<-glm(SF~sex*ldose, family=binomial)
> plot(numdead, ldose)
```

Then R will respond by plotting the graph;

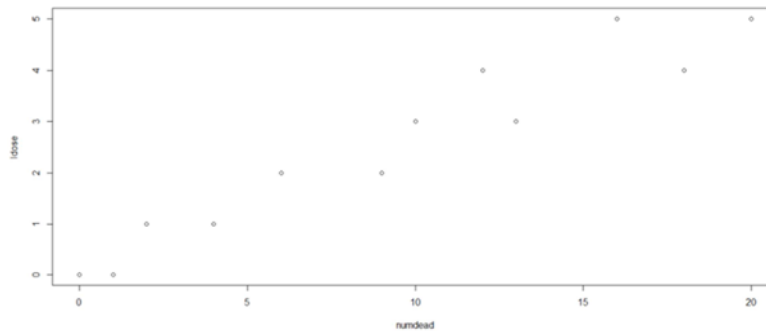


Figure 1. Showing the Plot of the Six Increasing Level of dose (i.e. 0 to 5) to the Number of Male (Points Above) and Female (Points Below) Moths that had Died. This Plot was Requested by the Last Command (i.e. "Plot (Numdead, Ldose)").

The issuance of the commands;

```
> summary(budworm)
```

```
> anova(budworm, test="Rao")
```

Will make R to give the result;

Call:

```
glm(formula = SF ~ sex * Ldose, family = binomial)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.39849	-0.32094	-0.07592	0.38220	1.10375

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-2.9935	0.5527	-5.416	6.09e-08 ***
sexM	0.1750	0.7783	0.225	0.822
Ldose	0.9060	0.1671	5.422	5.89e-08 ***
sexM:Ldose	0.3529	0.2700	1.307	0.191

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
(Dispersion parameter for binomial family taken to be 1)

Null deviance: 124.8756 on 11 degrees of freedom
Residual deviance: 4.9937 on 8 degrees of freedom

AIC: 43.104

Number of Fisher Scoring iterations: 4

Then the command;

```
> anova(budworm, test="Rao")
```

Gives the result;

Analysis of Deviance Table

Model: binomial, link: logit

Response: SF

Terms added sequentially (first to last)

	Df	Deviance	Resid.Df	Resid.Dev	Rao	Pr(>Chi)
NULL	11	124.876				
sex	1	6.077	10	118.799	6.051	0.0139 *
ldose	1	112.042	9	6.757	95.834	<2e-16 ***
sex:ldose	1	1.763	8	4.994	1.751	0.1858

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

The command;

```
> summary(glm(SF~sex+ldose,family=binomial))
```

Will then make R to give the result;

Call:

```
glm(formula = SF ~ sex + Ldose, family = binomial)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.10540	-0.65343	-0.02225	0.48471	1.42944

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-3.4732	0.4685	-7.413	1.23e-13 ***
sexM	1.1007	0.3558	3.093	0.00198 **
Ldose	1.0642	0.1311	8.119	4.70e-16 ***

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
(Dispersion parameter for binomial family taken to be 1)

Null deviance: 124.8756 on 11 degrees of freedom

Residual deviance: 6.7571 on 9 degrees of freedom

AIC: 42.867

Number of Fisher Scoring iterations: 4

3.3. When the Organisms are Inspected for Information on Toxicity

Here, the organisms (i.e. experimental units) or simply "units" are assembled and samples (e.g. blood and urine samples) are taken from each of them and used to estimate the quantities of the toxic elements available in each of the "sampled" organism. An array of quantities in which columns are allocated to toxic elements and rows, allocated to cases (i.e. units) is the matrix X, in the system of equation (3). The matrix of the response variables Y is usually unknown at the beginning, but with this technique, the matrix X will be used to data-mine some "hidden" information about Y. Such "data-mined" information on Y is either BRV-typed or count data typed depending on the quantity of hidden information that can be accessed through this technique. If universal "tolerance limits" exist for the units with respect to the toxic elements then a count data type Y is achievable otherwise (i.e. if they exist with respect to some or no toxic element), a BRV type Y is achievable.

3.4. Determination of Matrix Y When Tolerance Limits Exist for all Toxic Elements

The unit of measurement of the quantity of toxic element is

either “ppm” or “ $\mu g / dL$ ”, in this work; we shall assume the unit is $\mu g / dL$ throughout. Each data entry (x_{ij} , $i = 1, \dots, n$, $j = 1, \dots, m$) in matrix X is compared with its corresponding tolerance limit (τ_i , $i = 1, 2, \dots, m$) column-wisely, and instances in which the data entries are greater than their respective tolerance limits are counted and entered as the response variable y_i , $i = 1, \dots, n$ for the corresponding experimental unit. Matrix Y will be “created” the moment we enter the last response variable y_n . Here, the response matrix Y will contain nonnegative integers alone (i.e. $y_i \in \mathbb{Z}$, $0 \leq y_i \leq n$, $i = 1, 2, \dots, n$). The following section (3.5) contains a vivid numerical illustration of this technique.

3.5. Numerical Illustration on Vulnerability to Toxicity When all Tolerance Limits Exist

Table 1. Showing an Extract of the Matrix X with the Respective Tolerance Limits (Below the Line).

V2	V3	V4	V5	V6	V7	V8	V9
19.64	5.95	0.95	0.74	0.22	0.99	45.37	2.36
18.45	3.77	0.62	0.50	0.12	0.80	23.05	2.19
...							
30.1	5.14	1.51	1.24	0.37	0.51	24.89	1.63
28.25	6.30	0.82	0.88	0.20	0.42	17.45	1.16
32.14	5.52	1.06	0.94	0.25	0.57	18.53	1.66

The data for this illustration was obtained through samples from artisans operating in some mechanic villages (along Abeokuta-Ibadan expressways) around Abeokuta metropolis. The toxic elements are eight in number (i.e. V2, V3, ..., V9), 118 cases (or units) are used, an extract of the data and their respective tolerance limits are as contained in table 1 below (with all entries measured in $\mu g / dL$);

The corresponding extract of matrix Y (contained in column I or “V1”) is denoted $Y^T = (4, 3, \dots, 4, 1)$. The matrices Y and X are “supplied” together to R, such that the matrix Y occupies the “V1” location and the tolerance limits are excluded. The resulting “data-frame” is named “dat1”. The R codes for this operation are as contained in the three commands below;

```
> library("rJava")
> library("glmulti")
> dat1<- read.DIF("clipboard", transpose=TRUE)
```

If the above three commands are immediately followed by;

```
> outdat1<-glmulti(V1~V2+V3+V4+V5+V6+V7+V8+V9,
  data=dat1, method="g", maxit=30)
```

Then the Genetic algorithmic process to carry-out iterations and identify the formula (model) that will best “fit” the “contents” of dat1 is automatically initialized. An “extract” of the immediate response from R is;

Initialization.

TASK: Genetic algorithm in the candidate set.

Initialization.

Algorithm started.

After 10 generations:

Best model:

V1~1+V5+V7+V8+V3:V2+V4:V2+V5:V4+V6:V2+V6:V5+V7:V2+V7:V6+V8:V2+V8:V3+V8:V4+V8:V5+V8:V7+V9:V2+V9:V3+V9:V4+V9:V6

Crit= 291.254155883838

Mean crit= 310.663748809868

Change in best IC: -9708.74584411616 / Change in mean IC: -9689.33625119013

After 20 generations:

Best model:

V1~1+V7+V3:V2+V5:V2+V5:V4+V7:V2+V7:V3+V7:V6+V8:V3+V8:V4+V8:V7+V9:V3+V9:V4+V9:V6

Crit= 284.714137298027

Mean crit= 306.254977037995

Change in best IC: -6.5400185858104 / Change in mean IC: -4.40877177187315

...

After 750 generations:

Best model:

V1~1+V2+V4+V7+V9+V5:V2+V6:V3+V6:V4+V7:V2+V7:V4+V7:V6+V8:V2+V8:V4+V8:V7+V9:V5

Crit= 273.867533645176

Mean crit= 279.11632696666

Improvements in best and average IC have been going on below the specified goals.

Algorithm is declared to have converged.

Completed.

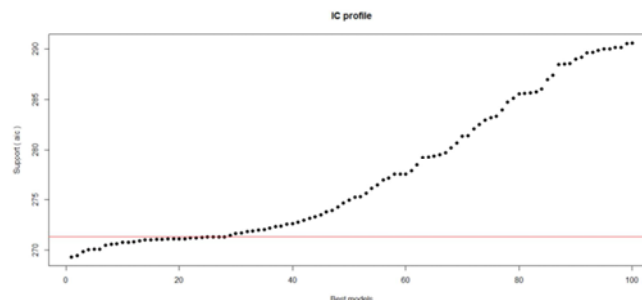


Figure 2. Showing that the Genetic Algorithmic Process Carried out 750 Iterations Before Identifying the Best Model to Fit the Data (dat1). The Plot of “Support(aic)” Versus “Best Models” Shows Where Lies the Critical Value Pictorially. Notice that the Critical (“crit”) and Mean Critical (“Mean Crit”)

Values were Continually Reducing as the Number of Iterations Increases until Convergence was Achieved After 750 Iterations with “Crit= 273.867533645176” and “Mean Crit= 279.11632696666”.

To further show that R’s choice of “Best model” is “reliable” and to carry-on, the researcher needs to give the codes below;

```
>out1<-glm(V1~1+V2+V4+V7+V9+V5:V2+V6:V3+V6:V4+V7:V2+V7:V4+V7:V6+
V8:V2+V8:V4+V8:V7+V9:V5, data=dat1, family=poisson)
> summary(out1)
```

The immediate response of R is as contained in the figure 3 below;

Call:

```
glm(formula = V1 ~ 1 + V2 + V4 + V7 + V9 + V5:V2 + V6:V3 + V6:V4 +
V7:V2 + V7:V4 + V7:V6 + V8:V2 + V8:V4 + V8:V7 + V9:V5, family = poisson,
data = dat1)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.69939	-0.29794	-0.03187	0.23211	1.13049

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-3.7725555	1.2709528	-2.968	0.00299 **
V2	0.0262955	0.0291362	0.903	0.36679
V4	1.9754260	0.9300434	2.124	0.03367 *
V7	4.6372329	2.2415605	2.069	0.03857 *
V9	0.5297197	0.3571043	1.483	0.13797
V2:V5	0.0271913	0.0170715	1.593	0.11121
V6:V3	0.2927339	0.1873984	1.562	0.11827
V4:V6	-4.6934531	2.1404095	-2.193	0.02832 *
V2:V7	-0.1021234	0.0492038	-2.076	0.03794 *
V4:V7	-1.6800130	0.8863794	-1.895	0.05804 .
V7:V6	8.7389122	5.2055934	1.679	0.09320 .
V2:V8	0.0008796	0.0008305	1.059	0.28954
V4:V8	0.0326369	0.0297313	1.098	0.27232
V7:V8	-0.0654351	0.0366379	-1.786	0.07410 .
V9:V5	-0.2016337	0.3462936	-0.582	0.56039

Signif. codes: 0 ‘***’ 0.001 ‘**’ 0.01 ‘*’ 0.05 ‘.’ 0.1 ‘ ’ 1

(Dispersion parameter for poisson family taken to be 1)

Null deviance: 93.529 on 117 degrees of freedom

Residual deviance: 26.106 on 103 degrees of freedom

AIC: 411.28

Number of Fisher Scoring iterations: 4

Figure 3. Showing the General Result from the Fitting of the Model (i.e. After Using R’s “Best Model” for Fitting “dat1”).

Now, besides the estimates of the coefficients, there are two noteworthy values, in figure 6, they are the “residual deviance” (26.106) and the “AIC” (411.28). These two values help “certify” it that our model certainly leads to the best fit. Further, if we

had chosen the “simplest” model (as it is usually done without the use of the “glmulti” function), then we would have supplied and receive (output) respectively, the content of figure 4 below;

```
> out2<-glm(V1~V2+V3+V4+V5+V6+V7+V8+V9, data=dat1, family=poisson)
> summary(out2)
OUTPUT:
Call:
glm(formula = V1 ~ V2 + V3 + V4 + V5 + V6 + V7 + V8 + V9, family = poisson,
    data = dat1)
Deviance Residuals:
    Min       1Q   Median       3Q      Max
-1.88896 -0.37321 -0.01591  0.36334  1.13949
Coefficients:
            Estimate Std. Error z value Pr(>|z|)
(Intercept) -1.318098  0.409665  -3.218  0.00129 **
V2           0.006861  0.005101   1.345  0.17866
V3           0.048752  0.048621   1.003  0.31601
V4           0.180747  0.195854   0.923  0.35608
V5           0.556853  0.361180   1.542  0.12313
V6           1.565954  1.219026   1.285  0.19893
V7          -0.220917  0.444327  -0.497  0.61905
V8           0.022283  0.010942   2.036  0.04170 *
V9           0.381777  0.141511   2.698  0.00698 **
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
(Dispersion parameter for poisson family taken to be 1)
Null deviance: 93.529  on 117  degrees of freedom
Residual deviance: 39.306  on 109  degrees of freedom
AIC: 412.48
```

Figure 4. Showing the Codes with the “Usual” Model (Without Glmulti) and the Corresponding Output.

Notice that;

1. The residual deviance that was 26.106 (with our best model) has now risen to 39.306 (with the usual model) and the AIC that was 411.28 has now risen to 412.48.
2. There is no “over-dispersion”, the evidence for this is contained in figure 5 below;

```
> summary(dat1$V1)
    Min. 1st Qu. Median Mean 3rd Qu. Max.
 0.000  2.000  3.000  3.517  5.000  8.000

> var(dat1$V1)
[1] 2.542445
```

Figure 5. Showing that There is no Over-dispersion Since the Variance of Y (i.e. 2.542445) is Less than Its Mean (i.e. 3.517) Which is an Unbiased Estimate of the Variance in Poisson Distribution.

Consequently, the fit for “dat1” is;

$$Y = -3.773 + 0.026*E_1 + 1.975*E_3 + 4.637*E_6 + 0.53*E_8 + 0.027*E_1*E_4 + 0.293*E_2*E_5 - 4.693*E_3*E_5 - 0.102*E_1*E_6 - 1.68*E_3*E_6 + 8.739*E_5*E_6 + 0.001*E_1*E_7 + 0.033*E_3*E_7 - 0.065*E_6*E_7 - 0.202*E_4*E_8 \quad (4)$$

Where E_i , $i = 1, 2, \dots, 8$, are the data entries per element per artisan and all other coefficients are approximated values of the coefficients in figure 4.

3.6. Further Diagnostic Checks on the Fit for Cases in Which all Tolerance Limits Exist

There are other diagnostic checks that corroborate the fact that equation (4) is the best fit for the data (dat1), some of them are;

1. The plot of Y against the residuals which gives the following (figure 3);

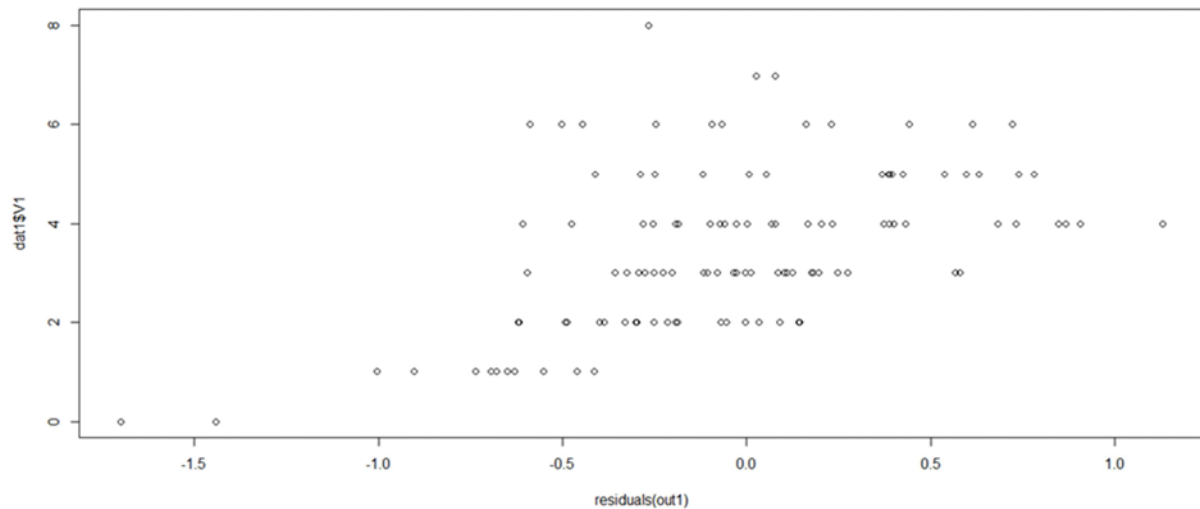


Figure 6. Showing That the Plot of Y Against the Residuals does not Connote any Relationship Between Them.

2. By comparing the AIC (for out1), in figure 6, with AIC (for out2), in figure 7, we can easily see that the fit for out1 is better than that for out2. The following pairs of statistics (figure 7) also testify to this fact;

```
> bic(out1)      > bic(out2)
[1] 452.8417      [1] 437.4181

> aicc(out1)     > aicc(out2)
[1] 415.9874      [1] 414.1486
```

Figure 7. Showing That the Fit for out1 is Better Than That From out2.

3. The “Hosmer and Lemeshow goodness of fit (GOF) test” for out1 (figure 8);

```
data: dat1$V1, fitted(out1)

X-squared = -4.3684, df = 8, p-value = 1
```

Figure 8. Showing the P-value Equals to 1, Which Means That the Corresponding Fit for out1 Cannot be Due to “Chance” (i.e. the Fit is Reliable) and the X-squared Statistic (i.e -4.3684) is also Good.

The vulnerability to toxicity of each artisan is a probability measure ($P(Y = y_i, i = 1, 2, \dots, n)$), its numerical value, for the entire data (dat1) will be obtained with the command “log10(fitted(out1))”. An extract of the probabilities is contained

in figure 9 below (i.e. by taken just four decimal places);

1	2	3	4	5	6	118
0.6562	0.3269	0.6433	0.8407	0.5641	0.5569	0.2546

Figure 9. Showing an Extract of the Vulnerability to Toxicity of the Artisans.

3.7. Determination of Matrix Y When Tolerance Limits Exist for Some or no Toxic Elements

The determination of the response matrix Y (i.e. a BRV) is through the matrix X which is used to data-mine it, using the following technique; the elements whose tolerance limits exist are “assumed” to be the “main” variables whilst all other variables are “assumed” to be “auxiliary”. In the data matrix X (i.e. in the equation 3), the first toxic element is actually “lead”. Although the human body does not possess any tolerance for lead (i.e. no matter how small the quantity of lead, it is still hazardous to man). However, Nriagu et al. (2008) helped determined the average lead in blood quantity of city children (aged 2 – 9 years) in Ibadan to be $(9.9 \pm 5.2 \mu\text{g} / \text{dL})$ which depicts that, a non-artisan “child” living in Ibadan and its environs could have as much as $(15.1 = 9.9 + 5.2 \mu\text{g} / \text{dL})$ in his/her blood. The mechanic villages from which the data in matrix X were obtained are on the two existing Abeokuta-Ibadan express-roads; hence the value $15.1 \mu\text{g} / \text{dL}$ is quite useful in the present work. If we assume that, as the children grow to be adults, they acquired more, say about $4.9 \mu\text{g} / \text{dL}$ “environmental” lead

into their body systems. Then a non-artisan adult in Ibadan and its environs is expected to have, on the average, $20 \mu\text{g} / \text{dL}$ of lead in his/her blood. Consequently, if an artisan has above this quantity (i.e. $20 \mu\text{g} / \text{dL}$) in his/her blood, we can “safely” assume that the additional quantity is due to occupational toxicity. The value $20 \mu\text{g} / \text{dL}$ was therefore utilized as the lead (i.e. the main element) toxicity limit for the artisans. By determining toxicity limits for the auxiliary variables (or by using their estimated population mean as toxicity limits), the auxiliary variable were used to “fine-tune” (in the sense that, if four or more of the auxiliary variables are above their toxicity limits, their corresponding y_i , $i=1,2,\dots,n$ that was formally 0 will become 1. Also y_i that was 1 before can become 0 if its main element is within +10 over its toxicity limit and only very few, say one of its auxiliary variable value is more than its toxicity limit) to obtain the concatenated matrix whose extract is;

V1	V2	V3	V4	V5	V6	V7	V8	V9
1	19.64	5.95	0.97	0.74	0.22	0.99	45.37	2.36
0	18.45	3.77	0.62	0.50	0.12	0.80	23.05	2.19
1	30.10	5.14	1.51	1.24	0.37	0.51	24.89	1.63
0	28.25	6.30	0.82	0.88	0.20	0.42	17.45	1.16
	20.0	5.52	1.06	0.94	0.2	0.57	18.53	1.66

Figure 10. Showing an Extract of the Concatenated Matrix (Y:X) Which is now the “Input” to R (Below the Line are the Toxicity Limits).

3.8. Numerical Illustration on Vulnerability to Toxicity When Tolerance Limits Exist for Some or no Toxic Elements

The corresponding data-frame is “dat3”, here; $Y = y_i = 0$

or 1 (i.e. 0 denotes that the vulnerability to toxicity is “relatively” low in this particular case when compared with the other cases in the data, whilst $y_i = 1$ denotes it is relatively high) we now proceed as before to obtain the best model that fits the data as (figure 12);

```
> outdat3<-
glmulti(V1~V2+V3+V4+V5+V6+V7+V8+V9,data=dat3,
method="g", family=binomial)
Initialization.
TASK: Genetic algorithm in the candidate set.
Initialization.
Algorithm started.
After 10 generations:
Best model:
V1~1+V2+V3+V5+V3:V2+V4:V2+V4:V3+V5:V2+V5:V3+
V5:V4+V6:V4+V6:V5+V7:V3+V7:V4+V7:V6+V8:V5+V8:
V6+V8:V7+V9:V3+V9:V7
Crit= 63.9154921762682
Mean crit= 797.229429038279
Change in best IC: -9936.08450782373 / Change in mean
IC: -9202.77057096172
After 580 generations:
Best model:
V1~1+V5+V6+V8+V9+V5:V4+V6:V2+V6:V4+V6:V5+V7:
V2+V7:V5+V7:V6+V8:V6+V9:V3+V9:V7+V9:V8
Crit= 47.7447083810335
Mean crit= 58.60998260074
Improvements in best and average IC have bebingo en
below the specified goals.
Algorithm is declared to have converged.
Completed.
```

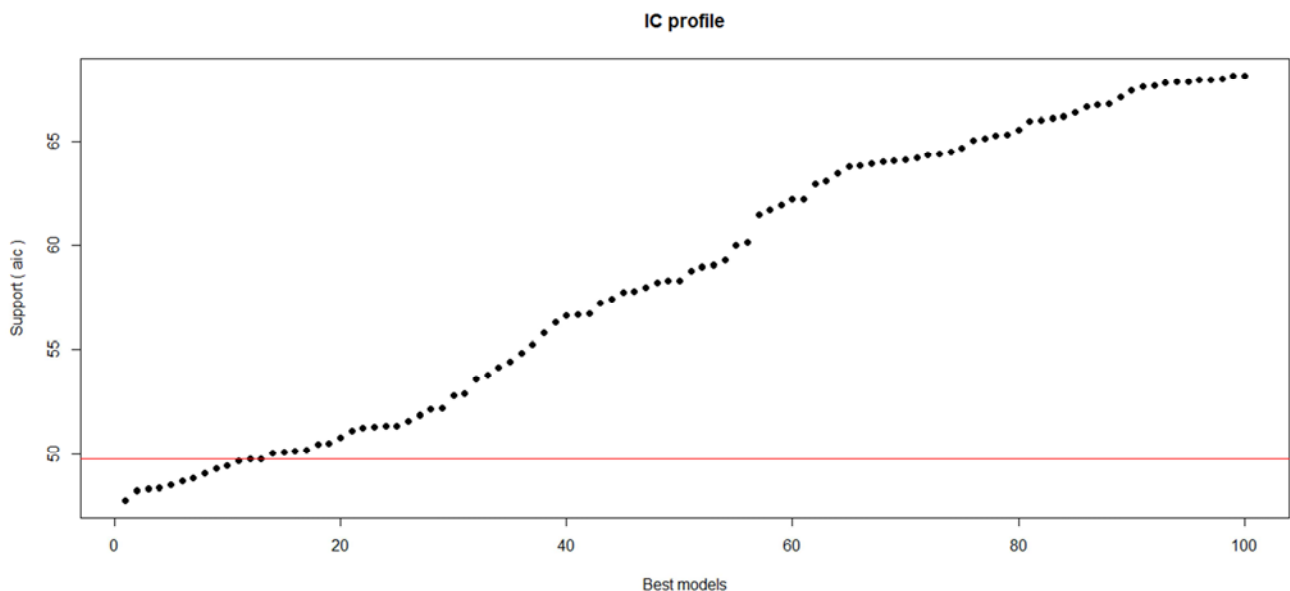


Figure 11. Showing the Extract of the Result When the R Command to Initialize the Iteration to Identify the Best Model (it was Achieved After 580 Iterations) was Given to R. The Plot of “Support(aic)” Versus “Best Models” Shows Where Lies the Critical Value Pictorially. As before, Notice That the Critical (“crit”) and Mean Critical (“Mean Crit”) Values were Continually Reducing as the Number of Iterations Increases Until Convergence was Achieved After 580 Iterations with “Crit= 47.7447083810335” and “Mean Crit= 58.60998260074”.

We now continue with a couple of commands in figure 12. That is;

```
>out3<-glm(V1~1+ V5+V6+V8+V9+V5:V4+V6:V2+V6:V4+V6:V5+V7:V2+V7:V5+
V7:V6+V8:V6+V9:V3+V9:V7+V9:V8, data=dat3, family=binomial)
> summary(out3)
```

Figure 12. Showing the Couple of Commands with Which the Model is Fitted and the Result Summarized Before the Display.

The result is as contained in figure 13 below;

```
Call:
glm(formula = V1 ~ 1 + V5 + V6 + V8 + V9 + V5:V4 + V6:V2 + V6:V4 +
V6:V5 + V7:V2 + V7:V5 + V7:V6 + V8:V6 + V9:V3 + V9:V7 + V9:V8,
family = binomial, data = dat3)

Deviance Residuals:
    Min       1Q   Median       3Q      Max
-2.217   0.000   0.000   0.000   1.523

Coefficients:
              Estimate Std. Error z value Pr(>|z|)
(Intercept)  1003.646   693.397   1.447   0.148
V5           -891.366   568.056  -1.569   0.117
V6          -8213.256  6026.54  -1.363   0.173
V8           -17.648   12.044  -1.465   0.143
V9            182.545   138.243   1.320   0.187
V5:V4         589.110   398.881   1.477   0.140
V6:V2          24.647    18.151   1.358   0.175
V6:V4        -1871.122  1258.687  -1.487   0.137
V5:V6        4450.705  3072.653   1.448   0.147
V2:V7         -6.856    5.311  -1.291   0.197
V5:V7       -1015.175   803.395  -1.264   0.206
V6:V7        7803.263  5974.693   1.306   0.192
V6:V8          90.718    60.156   1.508   0.132
V9:V3           2.004     1.486   1.348   0.178
V9:V7        -333.784   252.223  -1.323   0.186
V8:V9           1.493     1.336   1.117   0.264

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 160.176 on 117 degrees of freedom
Residual deviance: 15.745 on 102 degrees of freedom
AIC: 47.745

Number of Fisher Scoring iterations: 16
```

Figure 13. Showing the Result of the Fit for “dat3”.

Consequently, the fit for “dat3” is;

$$Y = 1003.646 - 891.366 * E_4 - 8213.256 * E_5 - 17.648 * E_7 + 182.545 * E_8 + 589.11 * E_3 * E_4 + 24.647 * E_1 * E_5 - 1871.122 * E_3 * E_5 + 4450.705 * E_4 * E_5 - 6.856 * E_1 * E_6 - 1015.175 * E_4 * E_6 + 7805.263 * E_5 * E_6 + 90.718 * E_5 * E_7 + 2.004 * E_2 * E_8 - 333.784 * E_6 * E_8 + 1.493 * E_7 * E_8 \quad (5)$$

Where E_i , $i = 1, 2, \dots, 8$, are the data entries per element per artisan and all other coefficients are approximated values of the coefficients in figure 13.

The vulnerability to toxicity of all the artisans are obtained together through the use of the following three commands (figure 14);

```
>l.hat=predict.glm(out3, dat3, se.fit=TRUE)
> c1=c(l.hat$fit-1.96*l.hat$se.fit, l.hat$fit+1.96*l.hat$se.fit)
> exp(c1)/(1+exp(c1))
EXTRACT OF OUTPUT
```

1	2	3	118
4.661908e-23	1.024383e-77	7.794027e-24	1.000000e+00

Figure 14. Showing the Three Commands with Which the Vulnerability to Toxicity are Requested for and an Extract of the Corresponding Output.

4. Conclusions

The following conclusions can be reached on the entire work, the issue of the command;

```
> anova(out1, test="Chisq")
```

will generate the following “analysis of deviance table” associated with the best fit (frame 11);

Analysis of Deviance Table

Model: poisson, link: log

Response: V1

Terms added sequentially (first to last)

	Df	Deviance	Resid.Df	Resid.Dev	Pr(>Chi)
NULL			117	93.529	
V2	1	1.3582	116	92.170	0.24384
V4	1	23.1972	115	68.973	1.462e-06 ***
V7	1	2.8963	114	66.077	0.08878.
V9	1	6.0721	113	60.005	0.01373 *
V2:V5	1	16.1289	112	43.876	5.918e-05 ***
V6:V3	1	2.4946	111	41.381	0.11424
V4:V6	1	0.6797	110	40.701	0.40970
V2:V7	1	1.5459	109	39.156	0.21374
V4:V7	1	4.5136	108	34.642	0.03363 *
V7:V6	1	1.6352	107	33.007	0.20098
V2:V8	1	2.7183	106	30.288	0.09920.
V4:V8	1	0.0027	105	30.286	0.95850
V7:V8	1	3.8414	104	26.444	0.05000.
V9:V5	1	0.3387	103	26.106	0.56056

 Signif. codes: 0 ‘***’ 0.001 ‘**’ 0.01 ‘*’ 0.05 ‘.’ 0.1 ‘ ’ 1

Figure 15. Showing the Analysis of Deviance Table, It has an Information on the Interaction of Elements; (1 and 4), (3 and 6), (1 and 7) and (6 and 7).

This gives the researcher some hints about some probable interacting toxic elements. Although the response, Y for the

case in which toxicity limits exist for some toxic elements has been coded with 0 and 1, but if it is coded as “FALSE” and “TRUE” (i.e. such that FALSE=0, TRUE=1), it will still work. These results ought to enhance the effectiveness of awareness campaigns informing artisans of the need to always put on their respective “safety” gadgets whenever they are at work. Artisans with high (i.e. TRUE) vulnerabilities to toxicity will know that they really have to exercise caution as much as possible. The results with respect to these coding technique are as contained in the following (figure 16, figure 17);

V1	V2	V3	V4	V5	V6	V7	V8	V9
TRUE	19.64	5.95	0.97	0.74	0.22	0.99	45.37	2.36
FALSE	18.45	3.77	0.62	0.50	0.12	0.80	23.05	2.19
TRUE	30.10	5.14	1.51	1.24	0.37	0.51	24.89	1.63
FALSE	28.25	6.30	0.82	0.88	0.20	0.42	17.45	1.16

Figure 16. Showing an Extract of “dat4” that was Supplied to R.

The dat4 was followed by the command;

```
outdat4<-
```

```
glmulti(V1~V2+V3+V4+V5+V6+V7+V8+V9,data=dat4,method="g", family=binomial)
```

which initiates the iterations to determine the best model that fits the data (dat4), an extract of the result of which is contained in figure 20 below;

Initialization.

TASK: Genetic algorithm in the candidate set.

Initialization.

Algorithm started.

After 10 generations:

Best model:

V1~1+V3+V4+V5+V6+V7+V3:V2+V4:V2+V5:V2+V5:V3+V5:V4+V6:V4+V6:V5+V7:V2+V7:V3+V7:V6+V8:V3+V8:V5+V8:V6+V8:V7+V9:V4+V9:V8

Crit= 63.9117692465579

Mean crit= 403.601512673779

Change in best IC: -9936.08823075344 / Change in mean IC: -9596.39848732622

After 810 generations:

Best model:

V1~1+V5+V8+V4:V3+V5:V4+V6:V2+V6:V4+V6:V5+V7:V2+V7:V3+V7:V4+V7:V5+V8:V7+V9:V6

Crit= 43.8845106069435

Mean crit= 63.4907240117881

Improvements in best and average IC have been going on below the specified goals.

Algorithm is declared to have converged.

Completed.

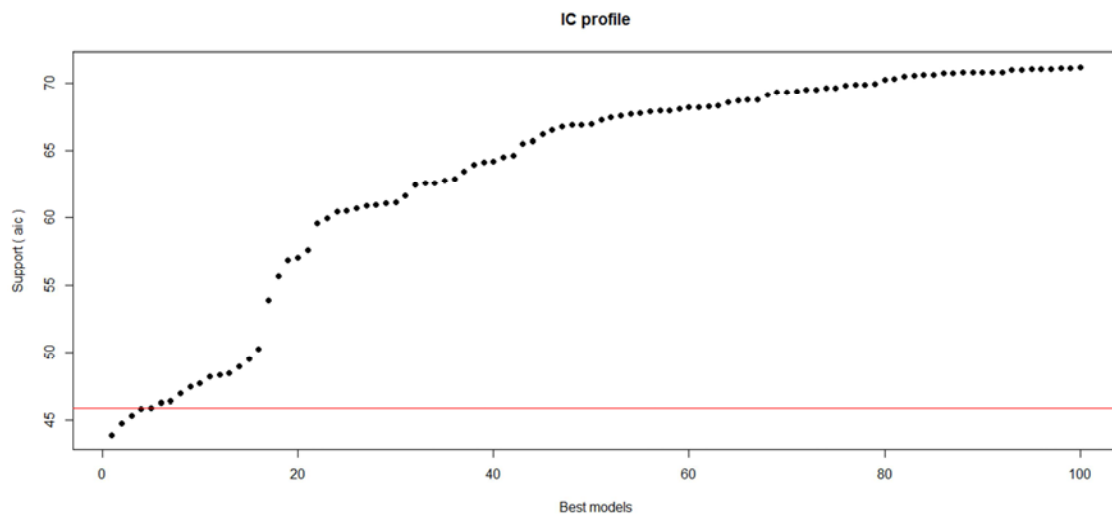


Figure 17. Showing the Extract of the Iteration Process that Identified the Best Model as well as the Corresponding Plot of the Support (aic) Versus Candidate “Best Models”.

With the couple of commands;

```
>out4<-glm(V1~1+V5+V8+V4:V3+V5:V4+V6:V2+V6:V4+V6:V5+V7:V2+V7:V3+V7:V4+V7:V5+V8:V7+V9:V6,
data=dat4, family=binomial)
> summary(out4)
```

The following result (i.e. frame 13) was obtained;

Call:

```
glm(formula = V1 ~ 1 + V5 + V8 + V4:V3 + V5:V4 + V6:V2 + V6:V4 +
    V6:V5 + V7:V2 + V7:V3 + V7:V4 + V7:V5 + V8:V7 + V9:V6, family = binomial,
    data = dat4)
```

Deviance Residuals:

Min	1Q	Median	3Q	Max
-1.884	0.000	0.000	0.000	1.504

Coefficients:

	Estimate	Std. Error	z value	Pr(> z)
(Intercept)	-704.050	661.871	-1.064	0.287
V5	-1078.197	969.022	-1.113	0.266
V8	40.369	37.161	1.086	0.277
V4:V3	-106.813	100.053	-1.068	0.286
V5:V4	1625.144	1496.963	1.086	0.278
V6:V2	8.927	9.155	0.975	0.330
V4:V6	-4297.377	3945.925	-1.089	0.276
V5:V6	3651.822	3317.727	1.101	0.271
V2:V7	2.515	2.042	1.232	0.218
V3:V7	252.605	234.955	1.075	0.282
V4:V7	786.453	737.168	1.067	0.286
V5:V7	-1508.985	1417.494	-1.065	0.287
V8:V7	-44.566	41.026	-1.086	0.277
V6:V9	424.902	395.336	1.075	0.282

(Dispersion parameter for binomial family taken to be 1)

Null deviance: 162.734 on 117 degrees of freedom

Residual deviance: 15.885 on 104 degrees of freedom

AIC: 43.885

Figure 18. Showing the Result of the Fit for dat4.

Consequently, the fit for dat4 is (equation 6);

$$\begin{aligned}
Y = & -704.05 - 1078.20 * E_4 + 40.37 * E_7 - 106.81 * E_2 * E_3 + 1625.14 * E_3 * E_4 + 8.93 * E_1 * E_5 \\
& - 4297.38 * E_3 * E_5 + 3651 * E_4 * E_5 + 2.52 * E_1 * E_6 + 252.61 * E_2 * E_6 + 786.45 * E_3 * E_6 \\
& - 1508.98 * E_4 * E_6 - 44.57 * E_6 * E_7 + 424.90 * E_5 * E_8
\end{aligned} \quad (6)$$

To predict the vulnerability to toxicity, the following set of commands is issued to R;

```

> pi.hat=predict.glm(out4, dat4, se.fit=TRUE)
> ci=c(pi.hat$fit-1.96*pi.hat$se.fit,
pi.hat$fit+1.96*pi.hat$se.fit)
> exp(ci)/(1+exp(ci)).

```

An extract of the result is contained in frame 14 ;

```

1          2          3          4          5
3.317545e-02 0.000000e+00 5.380727e-19 9.002411e-195 6.828425e-02
...

```

Figure 19. An Extract of the Result on Predicted Vulnerability to Toxicity.

Any of the approaches; depending on the type of data the researcher has (i.e. BRV or count data), could be adopted for any survey work on vulnerability to toxicity.

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